

In vivo genome engineering with CRISPR-Cas9

These publications demonstrate the application of CRISPR-Cas9 genome engineering techniques for target gene knockout, chromosomal deletions, precise knock-in, or gene correction in a variety of animal models.

Reference	Species/Cells	Target	Type of editing	Cas9 and guide RNA format	Delivery method	Link
Paix <i>et al. Nucl. Acids Res.</i> 2016	<i>Caenorhabditis elegans</i>	<i>dpy-10, gtbp-1, meg-3</i>	HDR/knock-in of reporter with ssODN or dsDNA, gene replacement	Cas9 protein and synthetic crRNA:tracrRNA (Dharmacon)	Microinjection	http://dx.doi.org/10.1093/nar/gkw502
Friedland <i>et al. Nat. Methods</i> 2013	<i>C. elegans</i>	<i>unc-119, dpy-13, klp-12, Y61A9LA.1</i>	Single gene knockout	Cas9 and sgRNA plasmids	Microinjection	http://dx.doi.org/10.1038/nmeth.2532
Wagner <i>et al. Nat. Methods</i> 2014	<i>Plasmodium falciparum</i>	<i>kahrp, eba-175</i>	HDR/knockin of reporter with plasmid donor	Cas9 and sgRNA plasmids	Electroporation	http://dx.doi.org/10.1038/nmeth.3063
Wang <i>et al. Cell Res.</i> 2013	<i>Bombyx mori</i> (silkworm) preblastoderm embryos	<i>BmBLOS2</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1038/cr.2013.146
Ma <i>et al. Sci. Rep.</i> 2014	<i>B. mori</i> (silkworm) eggs	<i>BmKu70</i>	Single gene knockout	Cas9 and sgRNA plasmids	Microinjection	http://dx.doi.org/10.1038/srep04489
Bassett <i>et al. Cell Rep.</i> 2013	<i>Drosophila</i> preblastoderm embryos	<i>yellow, white</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1016/j.celrep.2013.06.020
Li <i>et al. Sci. Rep.</i> 2015	<i>Papilio xhutus</i> (butterfly) eggs	<i>Abd-B, ebony, frizzled</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1038/ncomms9212
Kistler <i>et al. Cell Rep.</i> 2015	<i>Aedes aegypti</i> (mosquito) preblastoderm embryos	<i>AAEL010779, AAEL004091, AAEL000926, AAEL014228, AAEL002575, AAEL013647, Aaeg-wtrw</i>	Single gene knockout, HDR/knockout with ssODN, chromosomal deletion with two sgRNAs, HDR/knock-in of reporter with plasmid donor	Cas9 protein or mRNA and sgRNA	Microinjection	http://dx.doi.org/10.1016/j.celrep.2015.03.009
Hammond <i>et al. Nat. Biotechnol.</i> 2016	<i>Anopheles gambiae</i> (mosquito) zygotes	<i>AGAP005958, AGAP007280, AGAP011377</i>	HDR/knock-in/knockout of reporter with plasmid donor	Cas9 and sgRNA plasmids	Microinjection	http://dx.doi.org/10.1038/nbt.3439
Chang <i>et al. Cell Res.</i> 2013	<i>Danio rerio</i> (zebrafish) zygotes	<i>etsrp, gata5, gata4</i>	Single gene knockout and HDR/knock-in with ssODN	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1038/cr.2013.45
Hwang <i>et al. Nat. Biotechnol.</i> 2013	<i>D. rerio</i> (zebrafish) zygotes	<i>fh, th1, apoea, rgs4, tph1a, drd3, gria3a, slc6a3, tial1, gsk3b</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1038/nbt.2501
Jao <i>et al. PNAS</i> 2013	<i>D. rerio</i> (zebrafish) zygotes	<i>egfp, tyr, gol, mitfa, ddx19</i>	Single and multiple gene(s) knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1073/pnas.1308335110
Ansai & Kinoshita. <i>Biol. Open</i> 2014	<i>Oryzias latipes</i> (Japanese medaka) zygotes	<i>park7</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1242/bio.20148177
Nakayama <i>et al. Genesis</i> 2013	<i>Xenopus</i> zygotes	<i>tyr, six3</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1002/dvg.22720
Abu-Bonsrah <i>et al. Sci. Rep.</i> 2016	<i>Galus galus</i> (chicken) embryos	<i>DGCR8</i>	Single gene knockout	Cas9 and sgRNA plasmids	Microinjection, electroporation	http://dx.doi.org/10.1038/srep34524
Yang <i>et al. Cell</i> 2013	Mouse zygotes	<i>Tet1-loxP + Tet2-loxP, Sox2-V5, Nanog-mCherry, Oct4-GFP</i>	HDR/knock-in of reporter or epitope tag with ssODN or dsDNA	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1016/j.cell.2013.08.022
Han <i>et al. RNA Biol.</i> 2014	Mouse zygotes	<i>Rian</i>	Chromosomal deletion with two sgRNAs	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.4161/rna.29624
Platt <i>et al. Cell</i> 2014	Mouse brain and lungs	<i>Rbfox3, Kras + p53 + Stk11</i>	Single gene knockout, Kras G12D gene conversion with donor DNA	Cas9 tg-mouse, AAV-sgRNA	Stereotatic injection, intranasal, intratracheal instillation	http://dx.doi.org/10.1016/j.cell.2014.09.014

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Aida <i>et al. Genome Biol.</i> 2015	Mouse zygotes	<i>Actb</i>	HDR/knock-in of reporter with plasmid donor	Cas9 protein and synthetic crRNA:tracrRNA	Microinjection	http://dx.doi.org/10.1186/s13059-015-0653-x
Chen <i>et al. Cell</i> 2015	Mouse non-small-cell lung cancer cell line	Genes driving tumor growth and metastasis	<i>In vivo</i> screening	Cas9 and sgRNA stable integration	Lentiviral particles	http://dx.doi.org/10.1016/j.cell.2015.02.038
Dow <i>et al. Nat. Biotechnol.</i> 2015	Mouse ES cells	<i>Apc, Trp53, chrM. 8</i> nongenic region	Inducible single and multiple genes knockout	Cas9 and sgRNA tg-mouse	ESC transfection, transgenic mouse produced by blastocyst injection	http://dx.doi.org/10.1038/nbt.3155
Lupiáñez <i>et al. Cell</i> 2015	Mouse ES cells	<i>Epha4</i> TAD	Chromosomal deletion/inversion with two sgRNAs	Cas9 and sgRNA plasmids	ESC transfection, transgenic mouse produced by tetraploid complementation	http://dx.doi.org/10.1016/j.cell.2015.04.004
Wu <i>et al. Cell Res.</i> 2015	Mouse spermatogonial stem cells (SSC)	<i>Crygc</i>	Single gene knockout, HDR/gene correction with ssODN	Cas9 and sgRNA plasmids	SSC electroporation, transgenic mouse produced by SSC transplantation	http://dx.doi.org/10.1038/cr.2014.160
Long <i>et al. Science</i> 2016	Mouse cardiac and skeletal muscle	<i>Dmd</i>	Exon skipping	Cas9 and sgRNA AAVs	Intramuscular, intraperitoneal and retro-orbital injection	http://dx.doi.org/10.1126/science.aad5725
Nelson <i>et al. Science</i> 2016	Mouse cardiac and skeletal muscle	<i>Dmd</i>	Exon deletion	Cas9 and sgRNA AAVs	Intramuscular, intraperitoneal and intravenous injection	http://dx.doi.org/10.1126/science.aad5143
Tabebordbar <i>et al. Science</i> 2016	Mouse cardiac and skeletal muscle	<i>Dmd</i>	Exon deletion	Cas9 and sgRNA AAVs	Intramuscular, intraperitoneal and intravenous injection	http://dx.doi.org/10.1126/science.aad5177
Monteis <i>et al. Mol. Ther.</i> 2016	Mouse brain	Human <i>HTT</i> in tg-mouse	Allele-specific single gene knockout	Cas9 and sgRNA AAVs	Stereotactic injection	http://dx.doi.org/10.1016/j.ymthe.2016.11.010
Mikuni <i>et al. Cell</i> 2016	Mouse brain	<i>MeCP2, Actb, Dcx, Rab11a, Fmrp, Arc, Prkca, Cacna1c, Ywhae, Camk2a, Camk2b</i>	HDR/knock-in of epitope tag with ssODN	Cas9 and sgRNA plasmids	<i>In utero</i> electroporation	http://dx.doi.org/10.1016/j.cell.2016.04.044
Hu <i>et al. Cell Res.</i> 2013	Rat zygotes	<i>Dusp6, Gata5</i>	Single gene knockout, chromosomal deletion with 2 sgRNAs	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1038/cr.2013.141
Kaneko <i>et al. Sci. Rep.</i> 2014	Rat zygotes	<i>Il2rg</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Electroporation	http://dx.doi.org/10.1038/srep06382
Wang <i>et al. Sci. Rep.</i> 2015	Chinese Bama miniature pig zygotes	<i>Npc1l1</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1038/srep08256
Wang <i>et al. Mol. Ther. Nucleic Acids</i> 2016	Porcine fetal fibroblasts	<i>LRRK2</i>	HDR/knock-in, point mutation with ssODN	Cas9 and sgRNA plasmids	Cell electroporation, transgenic pig produced by somatic cell nuclear transfer	http://dx.doi.org/10.1038/mtna.2016.101
Wang <i>et al. Sci. Rep.</i> 2016	Sheep zygotes	<i>MSTN, ASIP, BCO2</i>	Single and multiple genes) knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1038/srep32271
Zou <i>et al. J. Mol. Cel. Biol.</i> 2015	Beagle dog zygotes	<i>MSTN</i>	Single gene knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1093/jmcb/mjv061
Niu <i>et al. Cell</i> 2014	Cynomolgus monkey zygotes	<i>Ppar-g + Rag1</i>	Multiple genes knockout	Cas9 mRNA and sgRNA (IVT)	Microinjection	http://dx.doi.org/10.1016/j.cell.2014.01.027

Abbreviations: IVT, *in vitro* transcription; HDR, homology-directed repair; ssODN, single-strand oligonucleotide donor; dsDNA, double-strand DNA; tg, transgenic; TAD, topologically associating domain; AAV, adeno-associated virus.

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