GE Healthcare

Publications with Dharmacon™ Edit-R™ CRISPR-Cas9 reagents

These publications demonstrate the application of CRISPR-Cas9 genome engineering techniques for target gene knockout or precise knockin using Dharmacon™ Edit-R™ CRISPR-Cas9 reagents or custom RNA synthesis.

2017

1. N. Maio, K. S. Kim, *et al.* A Single Adaptable Cochaperone-Scaffold Complex Delivers Nascent Iron-Sulfur Clusters to Mammalian Respiratory Chain Complexes I-III. *Cell Metab.* **25**, 945-953, e6 (2017). doi: 10.1016/j.cmet.2017.03.010
2. L. J. Rupp, K. Schumann K., *et al.* CRISPR/Cas9-mediated PD-1 disruption enhances anti-tumor efficacy of human chimeric antigen receptor T cells. *Sci Rep.* **7**, 737 (2017). doi:10.1038/s41598-017-00462-8
3. X. M. van Wijk, S. Döhrmann, *et al.* Whole-Genome Sequencing of Invasion-Resistant Cells Identifies Laminin α2 as a Host Factor for Bacterial Invasion. *MBio.* **8**, e02128-16 (2017). doi: 10.1128/mBio.02128-16

2016

1. K. He, E. Chou, *et al.* Conjugation and evaluation of triazole-linked single guide RNA for CRISPR-Cas9 gene editing. *ChemBioChem.* **17**, 1809–1812 (2016). doi:10.1002/cbic.201600320
2. R. Eggenschwiler, M. Moslem, *et al.* Improved bi-allelic modification of a transcriptionally silent locus in patient-derived iPSC by Cas9 nickase. *Sci Rep.* **6**, 38198 (2016). doi: 10.1038/srep38198
3. J. F. Hultquist, K. Schumann, *et al.* A Cas9 Ribonucleoprotein Platform for Functional Genetic Studies of HIV-Host Interactions in Primary Human T Cells. *Cell Reports* **17**, 1438–1452 (2016). doi:/10.1016/j.celrep.2016.09.080
4. M. L. Kelley, Ž. Strezoska, *et al.* Versatility of chemically synthesized guide RNAs for CRISPR-Cas9 genome editing. *J. Biotechnol.* **233**, 74–83 (2016). doi:10.1016/j.jbiotec.2016.06.011
5. J. McCaffrey, J. Sibert, *et al.* CRISPR-CAS9 D10A nickase target-specific fluorescent labeling of double strand DNA for whole genome mapping and structural variation analysis. *Nucleic Acids Res.* **44**, e11 (2016). doi:10.1093/nar/gkv878
6. A. Paix, H. Schmidt, *et al.* Cas9-assisted recombineering in C. elegans: genome editing using in vivo assembly of linear DNAs. *Nucleic Acids Res.* **44**, e128 (2016). doi:10.1093/nar/gkw502
7. J. Tan, S. E. Martin. Validation of Synthetic CRISPR Reagents as a Tool for Arrayed Functional Genomic Screening. *PLOS ONE* **11**, e0168968 (2016). doi: 10.1371/journal.pone.0168968

2015

1. E. M. Anderson, A. Haupt, *et al.* Systematic analysis of CRISPR-Cas9 mismatch tolerance reveals low levels of off-target activity. *J. Biotechnol.* **211**, 56-65 (2015). doi:10.1016/j.jbiotec.2015.06.427
2. R. Barrangou, A. Birmingham, *et al.* Advances in CRISPR-Cas9 genome engineering: lessons learned from RNA interference. *Nucleic Acids Res.* **43**, 3407–3419 (2015). doi:10.1093/nar/gkv226
3. H. Ogiwara, M. Sasaki, *et al.* Targeting p300 addiction in CBP-deficient cancers causes synthetic lethality by apoptotic cell death due to abrogation of MYC expression. *Cancer Discov.* **6**, 430-445 (2015). doi:10.1158/2159-8290.CD-15-0754
4. S. Opp, D. A. S. A. Vieira, *et al.* MxB Is Not Responsible for the Blocking of HIV-1 Infection Observed in Alpha Interferon-Treated Cells. *J. Virol.* **90**, 3056-3064 (2015). doi:10.1128/JVI.03146-15
5. A. Paix, A. Folkmann, *et al.* High efficiency, homology-directed genome editing in Caenorhabditis elegans using CRISPR-Cas9 Ribonucleoprotein complexes. *Genetics* **201**, 47-54 (2015) doi:10.1534/genetics.115.179382/-/DC1
6. G. Sivan, P. Ormanoglu, *et al.* Identification of Restriction Factors by Human Genome-Wide RNA Interference Screening of Viral Host Range Mutants Exemplified by Discovery of SAMD9 and WDR6 as Inhibitors of the Vaccinia Virus K1L-C7L- Mutant. *MBio.* 6, e01122 (2015). doi:10.1128/mBio.01122-15
7. W. Deng, X. Shi, *et al.* CASFISH: CRISPR/Cas9-mediated in situ labeling of genomic loci in fixed cells. Proc Natl Acad Sci U S A. 112, 11870–11875 (2015). doi:10.1073/pnas.1515692112

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